



MMP-13

Thermo Scientific MS-825P
Immunohistochemistry Protocol for Formalin Fixed Paraffin Embedded Tissue
Cut sections at 3 microns and bake overnight at 60°C

DAY 1

1. Bake slides at **60°C for 30 minutes** prior to starting IHC
2. Deparaffinize slides in xylenes for 5min each and rehydrate through graded alcohols (100% - 70% EtOH for 5 min each)
3. Wash in deionized water for 5 min each.
4. Enzymatic Antigen Retrieval: Perform antigen retrieval using **Hyaluronidase** (*Sigma H3506*)
See Hyaluronidase Digest in notes below.
5. Incubate slides **at 37°C** in a water bath **for 10 minutes**.
6. Rinse the slides in deionized water.
7. Outline each section with a **PAP pen**.
8. Quench endogenous peroxidase in **DAKO Endogenous Blocking Reagent for 30 min** (*Dako S2003*)
9. Rinse thrice with deionized water, then once in 1X PBST.
10. Block with **5% Normal Horse Serum for 30 minutes**. (*Vectastain Elite Mouse IgG PK-6102*).
11. Block non-specific binding sites **with the BEAT Blocking Solutions** (*Invitrogen cat# 50-300*) Add 2 drops of blocking solution **1A** to each section and **incubate for 30 min**.
12. Rinse the sections in deionized water.
13. Then rinse the sections **twice** in 1X PBST **for 3 minutes each**.
14. Add 2 drops of blocking solution **1B** to each section and **incubate for 10 min**.
15. Rinse the sections in deionized water.
16. Then rinse the sections **twice** in 1X PBST **for 3 minutes each**.
17. Drain and incubate **overnight at 4°C** with a **1:200** dilution of MMP-13 primary antibody (*Thermo Scientific MS-825P*) for **2-3 month old adult sections**.
18. Prepare the primary Ab in **2% Normal Horse Serum** (*Vectastain Elite Mouse IgG PK-6102*).
19. Use a **1:400** concentration for **E18.5 embryo sections**.
20. Negative Control slides need to be incubated with **2% Normal Horse Serum** only.



DAY 2

1. Let slides warm up to room temperature, then wash 5 times with 1X PBST for 5 min each.
2. Incubate the sections with **Vectastain Biotinylated Horse Anti-mouse secondary antibody** (*Vectastain Elite Mouse IgG PK-6102*) for **30 minutes**.
3. Reconstitute the **Vectastain ABC reagent** (*Vectastain Elite Mouse IgG PK-6102*) and incubate at **room temperature for 30 min**.
4. Wash 5 times with 1X PBST for 5 min each.
5. Incubate the sections with **Vectastain ABC reagent** for **30 minutes**.
6. Wash the slides well with 4 changes of 1X PBST, followed by 2 changes of deionized water
7. Detect color with **Vector DAB Impact** (*Vector SK-4105*) for a few minutes (check under the microscope)
8. Stop the color reaction with deionized water.
9. Counterstain the sections with **Hematoxylin** (*Zymed Cat # 93-3943*) for 5 minutes.
10. Rinse with tap water
11. Place slides in 1X PBS for 1-3 minutes.
12. Rinse with distilled water.
13. Dehydrate through 2 changes of 95% ethanol and 2 changes of 100% ethanol.
14. Clear in 3 changes of Xylene and then mount with Cytoseal.

¶ Notes-

Hyaluronidase Digest:

Dissolve 25mg of Hyaluronidase obtained from bovine testes (*Sigma Cat # H3506-1G*) in 250 ml of 1X PBS solution pre-heated to 37°C. Add the entire solution into a stain bucket. Place the slides in a slide rack and slowly immerse the rack into the warm Hyaluronidase solution. Cover the stain bucket with its lid and place in 37°C water bath for 10 minutes.

Vectastain Elite Mouse IgG Kit (PK-6102):

1:20 PBST-BSA can be used as the diluent buffer for the blocking serum and secondary antibodies. Follow the instructions written on the kit data sheet.

¶ Evidence of positive staining:-

Expression should be localized to the hypertrophic zones of the Growth plate. Articular Cartilage chondrocytes are **negative** for MMP-13 in wild type sections.