



Tak1 (Total)

Santacruz cat# sc7162

Immunohistochemistry Protocol for Formalin Fixed Paraffin Embedded Tissue

Cut sections at 3 microns and bake overnight at 60°C

DAY 1

1. Bake slides at **60°C for 30 minutes** prior to starting IHC
2. Deparaffinize slides in xylenes for 5 min each and rehydrate through graded alcohols (100% - 70% EtOH for 5 min each)
3. Wash in deionized water for 5 min each.
4. Enzymatic Antigen Retrieval – Perform antigen retrieval using **Hyaluronidase** (*Sigma H-3506*) Incubate slides at **37°C** in a water bath **for 10 minutes**.
5. See Hyaluronidase Digest in notes below.
6. Rinse the slides in deionized water.
7. Outline each section with a **PAP pen**.
8. Quench endogenous peroxidase in **DAKO Endogenous Blocking Reagent for 30 min** (*Dako S2003*)
9. Rinse thrice with deionized water, then once in 1X PBST.
10. Block non-specific binding sites with **the diluted Vectastain Normal Goat Serum** (*Vectastain Elite Rabbit Kit PK-6101*) – **incubate for 30 min**.
11. Do **NOT** rinse the serum off after the incubation period.
12. Drain and incubate **overnight at 4°C** with a **1:50** dilution of **Tak1 primary antibody** (*Santacruz cat# sc7162*) for **2-3 month old adult sections**.
13. Prepare primary Ab in **2% Normal Goat Serum**.
14. Negative Control slides need to be incubated with **2% Normal Goat Serum** only.



DAY 2

1. Let slides warm up to room temperature, then wash 5 times with 1X PBST for 5 min each.
2. Incubate the sections with **Vectastain Biotinylated Goat Anti-Rabbit secondary antibody** (*Vectastain Elite Rabbit Kit PK-6101*) for **30 minutes**.
3. Reconstitute the **Vectastain ABC reagent** (*Vectastain Elite Rabbit Kit PK-6101*) and incubate at **room temperature for 30 min**.
4. Wash 5 times with 1X PBST for 5 min each.
5. Incubate the sections with **Vectastain ABC reagent** for **30 minutes**.
6. Wash the slides well with 4 changes of 1X PBST, followed by 2 changes of deionized water
7. Detect color with **Vector DAB Impact** (*Vector SK-4105*) for a few minutes (check under the microscope)
8. Stop the color reaction with deionized water.
9. Counterstain the sections with **Hematoxylin** (*Zymed Cat # 93-3943*) for 5 minutes.
10. Rinse with tap water
11. Place slides in 1X PBS for 1-3 minutes.
12. Rinse with distilled water.
13. Dehydrate through 2 changes of 95% ethanol and 2 changes of 100% ethanol.
14. Clear in 3 changes of Xylene and then mount with Cytoseal.

¶ Buffers:-

1X PBST

Dilute 10 X PBS to 1X with deionized water.

Add Tween-20 to produce a final concentration of 0.01% Tween in 1X PBS.

Hyaluronidase Digest:

Dissolve 25mg of Hyaluronidase obtained from bovine testes (*Sigma Cat# H3506-1G*) in 250 ml of 1X PBS solution pre-heated to 37°C. Add the entire solution into a stain bucket. Place the slides in a slide rack and slowly immerse the rack into the warm Hyaluronidase solution. Cover the stain bucket with its lid and place in 37°C water bath for 10 minutes.

Vectastain Elite Rabbit IgG Kit (PK-6101):

1:20 PBST-BSA can be used as the diluent buffer for the blocking serum and secondary antibodies. Follow the instructions written on the kit data sheet.

¶ **Evidence of positive staining:-**

Expression should be localized to the hypertrophic zones of the Growth plate.
Articular Cartilage chondrocytes are **negative** for Tak1.

Protocol standardized on mouse tissue on 4/15/11 by Ashish Thomas, M.S.