STANDARD OPERATING PROCEDURE: Detection of Human Cytokine Response by Elispot in Human peripheral blood mononuclear cells following stimulation with antigen.

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1. Purpose:

The response of peripheral blood lymphocytes to antigen stimulation is measured by the capture of secrete cytokines with specific monoclonal antibodies bound to nitrocellulose filter microtiter plates by Elispot.

2. Principle:

The frequency of cytokine secreting cells in a lymphocyte population can be enumerated in Elispot. Frequencies are most often determined following antigenic stimulation in vivo or in vitro with antigen. A single anti-cytokine antibody diluted in PBS is allowed to bind to the wells of a polyvinylidene difluoride (PVDF) Elispot microtiter plate for a minimum of one hour at room temperature. The plates are washed with RPMI medium containing 8% fetal bovine serum/antibiotics/antimycotics before adding peripheral blood mononuclear cells (PBMC) at 1 to 5 X 10^5 cells per well and allowing antigen presenting cells to adhere to the well for 2 hours at 37° C. Either the non-adherent cells are removed (by washing 3X with RPMI with 8% serum) and CD4+ or CD8+ cells are selected to add back to the well or endogenous T cells from the PBMC are incubated with antigen/peptide for 24 hours to 48 hours. After incubation at 37° C the stimulated cells are removed and the plates are washed extensively with PBS-Tween. Biotinylated anti-cytokine antibody is added to the washed wells and incubated for 2 hours at room temperature. A streptavidin-alkaline phosphatase reagent is added for 30 minutes to 2 hours to detect biotinylated anti-cytokine antibody. Substrate is added to develop spots before reading on a CTL Elispot Analyzer with ImmunoSpot software.

3. Materials and Reagents:

- 96 well PVDF Elispot plates, Millipore, Catalog # MSIPN4W50
- Human peripheral blood mononuclear cells
- Specific anti-cytokine antibody or monoclonal antibody for coating (See Appendix A for specifics)
- Recall Antigens (See Appendix B for specifics)
- Paired biotinylated detection antibody (See Appendix A for specifics)
- Phosphate buffered saline (Dulbecco’s), Gibco Catalog # 14190-144
- Fetal Bovine Serum, Hyclone, Catalog # SH30396.03
- Bovine Serum Albumin (BSA), Bovuminar® Reagent Pure pH 7.0, Intergen, Cat # 3160-60
- Digital 20-200µL Multichannel Pipettor, VWR, Catalog # 40000-294
- Finnpipette 5-50µL single channel pipet, Thermo Electron, Cat # 4600220
- Finnpipette 1-10µL single channel pipet, Thermo Electron, Cat # 4600180
- Finnpipette 30-300µL single channel pipet, Thermo Electron, Cat # 4600240
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Sterile 20-200µL disposable pipet tips, Eppendorf, Catalog # 2249193-8
Sterile 30-300µL disposable pipet tips, Eppendorf, Catalog # 2249194-6
Sterile 0.1-20µL disposable pipet tips, Eppendorf, Catalog # 2249113-0
1 mL sterile disposable serological pipets, Costar, Catalog # 4012
5 mL sterile disposable serological pipets, Costar, Catalog # 4051
10 mL sterile disposable serological pipets, Costar, Catalog # 4101
25 mL sterile disposable serological pipets, Costar, Catalog # 4251

Pipet Aid, Drummond, Catalog # 4-000-101

Tween 20 detergent, Sigma, catalog # P1379
15 mL sterile polypropylene conical tubes, Falcon, Catalog # 35-2096
50 mL sterile polypropylene conical tubes, Falcon, Catalog # 35-2070
12x75mm culture tubes with closures, VWR Cat # 60818-500

Sterile reagent reservoirs, Costar, Catalog # 4871
37 °C incubator with 5% CO₂, Binder, Model # 9040-0030

Microplate washer, Biotek, Model ELx405 Select CW

CTL reader, Cellular Technology, LTD, Model ImmunoSpot® Series 3A Analyzer

Streptavidin-alkaline phosphatase conjugate, Jackson, Catalog # 016-050-084

Vector AP substrate kit, Catalog # SK-5300

Household bleach for decontamination, VWR, Catalog # 37001-056

Biological safety cabinet, Class II, Baker, Model # SG403

10X phosphate buffered saline (Dulbecco’s), Gibco, Catalog # 14200-075
5 mL syringe, VWR, Catalog # BD301603
10 mL syringe, VWR, Catalog # BD301604

Filter for syringe, Corning, Catalog #431215

Antibiotic/Antimycotic 100X Sigma Cat #A5955

ImmunoSpot® Software, Cellular Technology, LTD

RPMI 1640 medium, Cellgro, Catalog # 10-040-CV

RPMI 1640 with 8% FBS/Antibiotics/Antimycotics (R8A²)

PBS- 0.1% Tween 20

PBS-0.1TWEEN-2% BSA

100mM Tris-HCl Buffer pH 8.2

10% hypochlorite solution

Distilled water
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3.1 Reagent preparation:

3.1.1 Coating buffer: Dulbecco’s Phosphate Buffered Saline: 8 g NaCl; 0.2 g KCl; 2.16 g Na₂HPO₄·7H₂O; 0.2 g KH₂PO₄; add dH₂O to 1 liter. pH 7.2 sterile filter or autoclave. Store at 4°C. Expires 30 days after opening.

3.1.2 10X Wash buffer: 10 X PBS: Dulbecco’s Phosphate Buffered Saline: 80 g NaCl; 2 g KCl; 21.6 g Na₂HPO₄·7H₂O; 2 g KH₂PO₄; add dH₂O to 1 liter. pH 7.2. Store at room temperature. Expires after 3 months if not sterile.

3.1.3 Wash buffer: Phosphate Buffered Saline/0.1% Tween-20: Add 500mL of 10 X PBS to carboy. Add 5.0 mL of Tween-20. QS to 5 liters with distilled water. pH 7.2. Expires within two weeks.

3.1.4 Assay Buffer: Dulbecco’s Phosphate Buffered Saline/0.1% Tween-20/2%BSA: Add 100mL of 10 X PBS to clean beaker. Add 1 mL of Tween-20. Add 20 g BSA. QS to 1 liter with distilled water. Mix for 15 minutes on stir plate. pH 7.2. Sterile filter using a 0.45 micron filter into a sterile bottle. Expires within 24 hours if not sterile.

3.1.5 Media: RPMI/8%FBS: In hood, remove 40 mL RPMI from bottle and place in another sterile tube. Add 40 mL FBS. Add 5 mL antibiotics/antimycotics Mix. Store at 4°C.

3.1.6 Substrate Buffer: 100mM Tris-HCl pH 8.2: Add 12.114 g Tris to beaker. QS to 1 liter with Di water and mix. pH to 8.2 using HCl dropwise. Sterile filter using a 0.45 micron filter into sterile bottle. Store at room temperature. Expires after 3 months.

4. Procedure:

4.1. Prepare fresh or frozen Peripheral blood mononuclear cells as described below:

4.1.1. PBMC are isolated from fresh blood using ficoll hypaque separation or BD CPT tubes and spun twice at 1000 rpm for 10 minutes to remove platelets.

4.2. Dilute the sterile anti-cytokine antibody or Mab to the optimal concentration into sterile Dulbecco’s PBS. (See appendix A for Ab/MAB concentrations). Add 35 µL per well being careful not to touch the pipet tips to the filter. Tap the plate to wet the wells and verify that the wells are entirely coated. Incubate the Elispot plate wells for 1 to 3 hours at room temperature or overnight at 4°C to coat the filter with antibody.

4.3. Add 100 µL R8A² to each well in the plate using a multichannel pipet and aspirate using a sterile aspirator being careful not to touch the pipet tips to the filter. Repeat two times.
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4.4. Add 50 µL of antigen diluted to the appropriate concentration in R8A² to the appropriate wells of the plate and then add 50 µL per well of the cell suspension in R8A² that has been gently and thoroughly mixed being careful not to touch the pipet tips to the filter (Reference Appendix B). Cover microtiter plate and place in 37°C incubator with 5 % CO₂ for 4 to 48 hours depending on the cytokine to be detected. Reference appendix C. Do not stack plates. Be careful not to tip or shake when handling. (For IL-4 and IL-5, the volume is 100 µL per well for a total volume of 200 µL)

4.5. Inspect plate visually to determine if any contamination is present. Wells with discolored media may be an indication of contamination.

4.6. Wash the plate on the microtiter plate washer three times with 250µL per well of PBS- 0.1% tween-20 soaking for 5 seconds between each wash. Repeat wash step.

4.7. Add 30 µL per well of the optimal concentration (Reference Appendix A) of the (sterile filtered) biotinylated detection antibody in PBS-0.1%Tween with 2% BSA being careful not to touch the pipet tips to the filter and incubate for 2 hours at room temperature. Wash 3 times as in step 4.6.

4.8. Add 30 µL per well of streptavidin-alkaline phosphatase at a 1:1000 dilution in PBS-0.1%Tween with 2% BSA and incubate at r.t. for 1 hour. Wash 3 times as in step 4.6.

4.9. Soak the plates in PBS-0.1%Tween for 1 hour at room temperature after removing the back of the Elispot plate. After 1 hour, dry plate off by blotting dry onto absorbent paper.

4.10. Develop by adding 100 µL per well being careful not to touch the pipet tips to the filter of Vector AP substrate kit III at room temperature in the dark from 3 to 120 minutes. Aspirate out the substrate using the plate washer or multichannel pipett being careful not to touch the pipet tips to the filter. Wash the plate on a microtiter plate washer as indicated in 4.6 using distilled water. Allow plates dry overnight.

4.11. After the plates have dried, read on CTL reader.
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Appendix A: Coating and Detection Antibody Concentration

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>Species</th>
<th>Coating Source</th>
<th>ELISpot Coat Concentration</th>
<th>Coating Clone</th>
<th>Detection Source</th>
<th>Detection concentration</th>
<th>Detection Clone</th>
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<tbody>
<tr>
<td>IFNγ</td>
<td>human</td>
<td>Mab tech 3420-3-1000</td>
<td>10µg/mL</td>
<td>1-D1K</td>
<td>Mab tech 3420-6-1000</td>
<td>1µg/mL</td>
<td>7-B6-1</td>
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<tr>
<td>IL-2</td>
<td>human</td>
<td>Mab tech 3440-3-1000</td>
<td>10µg/mL</td>
<td>IL2-I</td>
<td>Mab tech 3440-6-1000</td>
<td>1µg/mL</td>
<td>IL2-II</td>
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<tr>
<td>IL-4</td>
<td>human</td>
<td>Mab tech 3410-3-18</td>
<td>15µg/mL</td>
<td>TL4-I (82.4)</td>
<td>Mab tech 3410-6-11</td>
<td>2µg/mL</td>
<td>IL4-II (12.1)</td>
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<tr>
<td>IL-5</td>
<td>human</td>
<td>Mab tech 3490-3-1000</td>
<td>10µg/mL</td>
<td>TRFK5</td>
<td>Mab tech 3490-6-1000</td>
<td>1µg/mL</td>
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NOTE: Alternative antibodies for IL-2 can be obtained from R&D Systems, Catalog number SEL202 and catalog number SEL202. Alternative antibodies for IL-5 can be obtained from BD Pharmingen, catalog #551085(coating and detection antibody included).
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Appendix B: Recall Antigen Concentrations

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<thead>
<tr>
<th>Recall Antigen</th>
<th>Source</th>
<th>Concentration</th>
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<tbody>
<tr>
<td>Tetanus Toxoid</td>
<td>Calbiochem</td>
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<tr>
<td>Inactivated Flu 4907151</td>
<td>Charles River SPAFAS</td>
<td>5µg/mL</td>
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<td>RSV</td>
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Appendix C: Incubation time

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<th>Cytokine</th>
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<tr>
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<td>IL-2</td>
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<tr>
<td>IL-4</td>
<td>40</td>
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<tr>
<td>IL-5</td>
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Revision History

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